THE EFFICIENCY OF BOVINE ELISA IN DETECTION OF THE Mycobacterium avium SUBSPECIES Paratuberculosis (MAP) INFECTION IN CAMEL (Camelus dromedaries) AT DIFFERENT AGES

Ahmed M Alluwaimi

Department of Microbiology and Parasitology, P O Box 35252, College of Veterinary Medicine and Animal Resources, King Faisal University, Al-Ahsa 31983, Saudi Arabia

ABSTRACT

Mycobacterium avium subspecies *Paratuberculosis* infection is chronic disease that infect young animals and remains undetected for long period. The immune responses to the disease is characterised by the cell mediated immunity at the early stage while antibody response dominates the late stage of the infection. Early detection of the disease is vital to prevent its transmission to the susceptible animals. ELISA was seen one the sensitive tests in detection of the disease. The application of bovine test in detecting the disease was sought to unravel its versatility in camel. 95 serum samples [2-3 years-old (3 samples), 4-6 years-old (21 samples), 7-9 years old (24 samples) and 10-15 years-old (47 samples)] were collected from dromedary camels. The analysis of serum samples with commercial ELISA indicated only 8 positive (8.4%) and one inconclusive samples. The positive samples were restricted to older ages (7-9 years-old and 10-15 years-old). Despite the emaciation of few animals, the postmortem revealed no significant changes in intestine. The results proved that bovine ELISA is feasible in detecting anti-MAP antibodies in camel. However, the limitation of the ELISA sensitivity in detecting the infection in young animals will leave wide range of infected young animals undetected. The results encourage the application of ELISA concurrent with PCR and/or faecal culture in study of MAP infection prevalence in camel in Saudi Arabia.

Key words: Bovine ELISA, camel, Mycobacterium avium subspecies Paratuberculosis

Mycobacterium avium subspecies *Paratuberculosis* (MAP) causes John's disease in domestic and wild ruminant like cattle, sheep, goats, deer, antelope and bison (Stabel, 1997) worldwide. In Saudi Arabia, John's disease was reported in sheep, goat, dairy cattle, and camel (Ahmed and Towfik, 1999; Gameel *et al*, 1994; Alluwaimi *et al*, 1999; Al Hajri and Alluwaimi, 2007).

Long incubation period is the main characteristic feature of MAP infection. Ingestion of faecal material, milk or colostrum is the main route of infection. Infected cattle shed low amount of bacteria during the subclinical stage. However, during the clinical stage the shaded organisms in faeces increase dramatically. At the clinical stage, infected animals manifest chronic diarrhoea, emaciation, decrease milk production and infertility (Stabel, 1997).

Immunodiagnostic techniques were applied to assure the early diagnosis of MAP infection. Early diagnosis of MAP infection represents one of the major obstacles in successful control of the disease (Valentin-Weignad, 2002). During the subclinical phase, the immune responses are dominated by cell-mediated immunity whereas, humoral immune responses prevail at the clinical phase (Coussens, 2001; Valentin-Weignad, 2002). Different versions of enzyme linked immunosorbent assay (ELISA) and molecular based techniques were introduced in the last decade to overcome the impediments of detecting the subclinical MAP infection. Nevertheless, the newly advanced techniques were not sensitive enough unless they were used in suitable combination to achieve clear cut diagnosis (Collins *et al*, 1993a, b; Collins, 1996; Paolicchi *et al*, 2003; Moss *et al*, 1991).

ELISA remains one of the most applied tests for the early detection of MAP infection. Overwhelming studies examined the sensitivity and specificity of ELISA in the detection of subclinical MAP infection (Nielsen and Toft, 2006; Collins, 2002; 2005). The sensitivity of ELISA in detecting the infection at the early stage of the infection is about 15%. ELISA sensitivity in moderately shedding animals is only 47-48%. However, it scores 88% in animals with clinical signs (Whitlock *et al*, 2000). Although, ELISA was

SEND REPRINT REQUEST TO AHMED M ALLUWAIMI email: alluwaimi@saudivms.org.sa

developed to detect anti-MAP antibodies in camelid in south America (Kramsky *et al*, 2000), there are no information on application of this newly introduced techniques and their versatility in detecting MAP infection in dromedary camels.

The main objective of this study was to assure the feasibility of bovine ELISA kit as a robust technique in detecting at the late subclinical and early clinical stages of MAP infection. The efficiency of ELISA in detecting MAP infection will assure stringent approaches in control of the disease in the country.

Materials and Methods

Pre and post partum examination

Animals were examined before slaughter for the emaciation and diarrhoea. The ileum and mesenteric lymph nodes were examined after the slaughter.

Serum samples

A total of 95 serum samples were collected from different ages of camel at Dammam abattoir. The samples were arranged in 4 groups as follows, 2-3 years-old (3 samples), 4-6 years-old (21 samples), 7-9 years old (24 samples) and 10-15 years old (47 samples).

ELISA for the detection of the anti-MAP antibodies

The camel serum samples were analysed for the presence of anti-MAP antibodies using ID Vet ELISA kit (ID VET, Montpellier, France). After preabsorption step with Mycobacterium phlei to remove potential cross-reacting antibodies, 100 µl of the previously neutralised samples and controls were transferred to the coated ELISA micoroplate and incubated for 45 minutes at room temperature. After thorough washing 100 µl of 1X conjugate was then added to each well and incubated for 30 minutes at room temperature. 100 µl of substrate solution was then added to each well after rewashing and incubation in dark for 15 minutes at room temperature. Then 100 µl of stop solution was added to each well. The plate was read by ELISA reader (Thermo Labsystem, Finland) at dual wavelength 450 and 620 nm.

Results

The analysis of serum samples with ELISA

The serum analysis with ELISA revealed 8 positive (8.4%) and one doubtful (1%). The positive samples were restricted to the old animals [6 samples of 10-15 years old (12.7%) and 2 samples of 7-9 years-old (8.3%)]. Table 1 refers to the distribution of serum samples analysis according to age.

The pre and postmortem examination

Only one of the few emaciated camels appeared to be ELISA positive. The postmortem examination of ileum and mesenteric lymph nodes of the emaciated and other camels failed to reveal any significant histological changes.

Discussion

This study examined the possibility of detecting anti-MAP infection by bovine ELISA kit. It is considered the first to attempt to test the feasibility of this approach in detecting the MAP infection in camel.

MAP infection is chronic debilitating disease. Animals are infected at their early stage of life (0-4 months). However, the development of the disease is subjected to various factors like infective dose and age of the animal. Hence, three groups of animals in regard to MAP infection can be classified as affected, infectious and infected (Nielsen and Toft, 2008). The affected animals are those that manifest clinical signs such as diarrhoea, reduction of milk production and chronic weight loss. Infectious animals on the other hand, comprise those animals that shed MAP and considered a source of transmission to the susceptible animals. The third group is the infected group which lies between the affected and the infectious groups. In the infected animals MAP persists in macrophages.

In relation to the above mentioned classification, the sensitivity of ELISA is greatly influenced by the type of the group. Therefore, animals shedding less than 10 colonies per tube are more likely to be seronegative, while shedding more than 70 colonies per tube are considered strong seropositive. Hence, ELISA sensitivity in cattle naturally infected with MAP could be as low as 15% in low shedding young animals, whereas in the moderately shedding animals the sensitivity could reach to 47-48%, while the heavily shedding animals the sensitivity is 88% (Whitlock *et al*, 2000).

In view of these facts the collection of the samples was concentrated mainly on the animals above 3 years because the application of ELISA was

Table 1. The total analysis of ELISA according to age of camels.

Age	Positive	Negative	Inconclusive	Total	Age (%)
2-3	0	2	1	3	3
4-6	0	21	0	21	22
7-9	2	22	0	24	25
10-15	6	41	0	47	50
Total	8	86	1	95	100
Total (%)	8.4	90.6	1	100	

most likely to reveal more positive samples in old animals. In consistence with this approach the results of this study are in accordance with the previous report (Al Hajri and Alluwiami, 2007). Furthermore, the results reflect the feasibility of the MAP antigen in the ELISA kit for detecting the disease in camel as well as the practicality of the conjugated antibodies to detect the camel anti-MAP antibodies. Nevertheless, limitation of ELISA in detecting the infection in young camels will leave wide range of young infected camels (below 4 years) undetected. Hence, concurrent application of polymerase chain reaction (PCR) with ELISA is essential in increasing the detection range of the infected animals.

In general, this preliminary study provides promising insight in utilising bovine ELISA in the study of the prevalence of the MAP infection in *dromedariy camel* in Saudi Arabia.

References

- Ahmed SA and Towfik A (1995). Johne's disease among sheep and goat of El-Qassiem area. 16th Annual Meeting of Saudi Biological Society.
- Alluwaimi AM, Hatem ME and Almousa JM (1999). The efficacy of gel immunodiffusion and faecal smear tests for diagnosis of ovine paratuberculosis in sheep in Saudi Arabia. The Egyptian Journal of Immunology 7:29-32.
- Al Hajri SM and Alluwaimi AM (2007). The efficiency of ELISA and PCR in detecting subclinical paratuberculosis in the Saudi dairy herds. Pakistan Journal of Biological Science 10:1906-1909.
- Collins MT, Scott J, Wells 2, Kristine R Petrini, James E, Collins Ronald D, Schultz 1 and Robert Whitlock H (2005). Evaluation of 5 antibody detection tests for diagnosis of bovine paratuberculosis. Clinical and Diagnostic Laboratory Immunology 12:685-692.
- Collins MT (2002). Interpretation of a commercial bovine paratuberculosis enzyme-linked immunosorbent assay by using Likelihood Ratios. Clinical and Diagnostic Laboratory Immunology 9:1367-13.
- Collins MT (1996). Diagnosis of paratuberculosis. Veterinary Clinics of North America Food Animal Practice 12:357-71.
- Collins DM Stephens DM and de Lisle GW (1993a). Comparison of polymerase chain reaction tests and faecal culture

for detecting *Mycobacterium paratuberculosis* in bovine faeces. Veterinary Microbiology 36:289-99.

- Collins MT, Buergelt AA, Hennager CD, Hietala SG, Jacobson SK, Whipple DL and Whitlock RH (1993b). Reproducibility of a commercial enzyme-linked immunosorbent assay for bovine paratuberculosis among 8 laboratories. Journal of Veterinary Diagnostic Investigation 5:52-5.
- Coussens PM (2001). *Mycobacterium paratuberculosis* and the bovine immune system. Animal Health Research 2:141-161.
- Gameel AA, Ali AS Razig SA, Brown J and Elhendi A (1994). A clinico-pathological study on spontaneous paratuberculosis in camels (*Camelus dromedarius*). Pakistan Veterinary Journal 14:15-19.
- Kramsky JA, Miller DS, Hope A, and Collins MT (2000). Modification of a bovine ELISA to detect camelid antibodies to *Mycobacterium paratuberculosis*. Veterinary Microbiology 77(3-4):333-7.
- Moss MT, Green EP, Tizard ML, Malik ZP and Hermon-Taylor J (1991). Specific detection of *Mycobacterium paratuberculosis* by DNA hybridisation with a fragment of the insertion element IS900. Gut 32:395-8.
- Nielsen SS and Toft N (2008). Ante mortem diagnosis of paratuberculosis: A review of accuracies of ELISA, interferon-gamma assay and faecal culture techniques. Veterinary Microbiology 129(3-4):217-35.
- Nielsen SS and N Toft (2006). Age-specific characteristics of elisa and fecal culture for purpose-specific testing for paratuberculosis. Journal of Dairy Science 89:569-579.
- Paolicchi FA, Zumarraga MJ, Gioffre A, Zamorano P, Morsella C, Verna A, Cataldi A, Alito A and Romano M (2003). Application of different methods for the diagnosis of paratuberculosis in a dairy cattle herd in Argentina. Journal of Veterinary Medicine B. 50:20-26.
- Stabel JR (1997). John's disease: A hidden threat. Journal of Dairy Science 81:283-288.
- Valentin-Weigand P (2002). Johne's disease: pathogenesis and problems related to diagnosis. In: In; Recent developments and perspectives in bovine medicine. Ed. By Kaske, M., Scholz, H. Holtershinken, M., pp. 48-57. XXII World Buiatrics Congress, 18-23 Aug, 2002. Hannover, Germany.s
- Whitlock RH, Wells SJ, Sweeney RW and Van Tiem J (2000). ELISA and faecal culture for paratuberculosis (John's disease): sensitivity and specificity of each method. Veterinary Microbiology 77:387-398.